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Isolation, Structure Elucidation, and Synthesis of Eudistomides A and B, Lipopeptides from a Fijian Ascidian *Eudistoma* sp.

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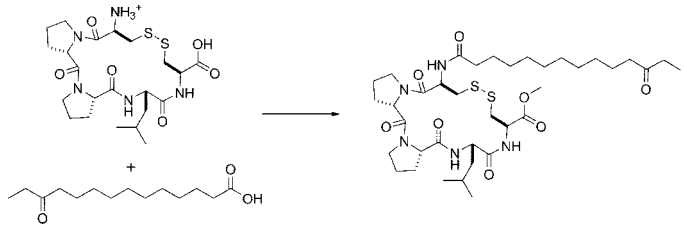
Received October 8, 2008



Eudistomides A (**1**) and B (**2**), two new cyclic peptides, were isolated from a Fijian ascidian *Eudistoma* sp. These five-residue cystine-linked cyclic peptides are flanked by a C-terminal methyl ester and a 12-oxo- or 12-hydroxy-tetradecanoyl moiety. The complete structures of the eudistomides were determined using a combination of spectroscopic and chemical methods. Chiral HPLC analysis revealed that all five amino acid residues in **1** and **2** had the L-configuration. Total synthesis of eudistomides A (**1**) and B (**2**) confirmed the proposed structures. Enantioselective lipase-catalyzed hydrolysis of a mixture of C-35 acetoxy epimers indicated a 35*R* absolute configuration for **2**.

Introduction

Ascidians are known to be a rich source of complex and structurally unique peptides such as ulithiacyclamides,[1−3] patellamides,[3−7] lissoclinamides,[7−9] and didemnins.[10−15] To date, however, peptides have not been reported from the genus *Eudistoma*. Previous investigations of *Eudistoma* have yielded eudistomins,[16−20] eudistomidins,[21−24] iejimalides,[25,26] and many alkaloids,[27−37] several of which are brominated.[33−37] As part of the continuing search for structurally unique secondary

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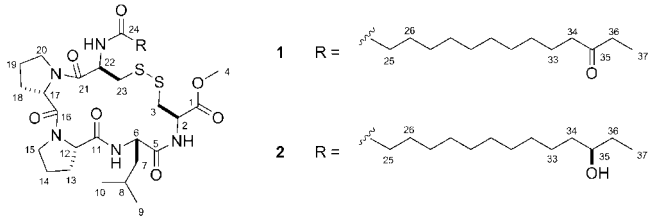
10.1021/jo8022582 CCC: $40.75
Published on Web 12/03/2008

metabolites from marine invertebrates, a detailed exploration of the morphologically distinct Fijian ascidian *Eudistoma* sp. was undertaken. The isolation, structure elucidation, and synthesis of two new *Eudistoma*-derived lipopeptides, eudistomides A (**1**) and B (**2**), are described herein.

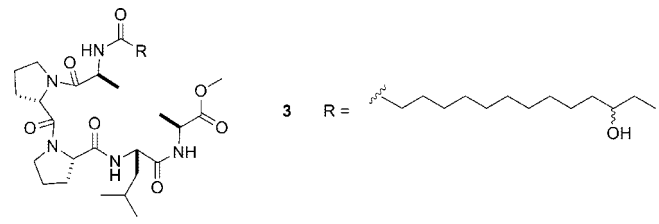
Results and Discussion



The specimen (FJ04-12-071) of *Eudistoma* sp. was lyophilized, ground to a fine powder, and exhaustively extracted with MeOH. The crude extract was subjected to an EtOAc/H_2O partition, and the EtOAc soluble material was separated on HP20SS resin using a step gradient of H_2O to acetone (10% steps, 11 fractions). The sixth (50/50 acetone/H_2O) and seventh (60/40 acetone/H_2O) fractions were combined and purified using several rounds of reversed-phase HPLC, resulting in the isolation of eudistomides A (**1**) and B (**2**).

Eudistomide A (**1**) showed an $[M + Na]^+$ ion at *m/z* 790.3897 in the HRESIMS, which is consistent with the molecular formula $C_{37}H_{61}N_5O_8S_2$ (calcd for $C_{37}H_{61}N_5O_8S_2Na$, 790.3859; Δ +4.8 ppm), and required 10 degrees of unsaturation. Initial evaluation of the ^1H and ^{13}C spectra suggested that **1** was a peptide (Table 1). The peptide nature of the molecule was further supported by the presence of three exchangeable NH signals at δ_H 6.08 (d, 10.5), 7.20 (d, 9.3), and 9.42 (d, 8.1) in the ^1H spectrum. The ^{13}C NMR spectrum showed six carbonyl signals (amides and/or esters) at δ_C 169.4, 170.1, 170.9, 171.5, 171.8, and 173.2. The presence of an ester group in **1** was confirmed by analysis of the IR spectrum which displayed a characteristic absorbance band at $\nu_{max} \approx 1740$ cm^{-1}. The ^1H and HMBC data corroborated the identity of the methyl ester; a methyl resonance at δ_H 3.73 (H-4) showed an HMBC correlation to a carbonyl at δ_C 170.9 (C-1). A carbon resonance at δ_C 212.1 (C-35) indicated the presence of a ketone in **1**. The planar structure of eudistomide A (**1**) was assigned after extensive one-dimensional (1D) and two-dimensional (2D) NMR studies. Analysis of the 1D TOCSY, COSY, HSQC, and HMBC data established the presence of five amino acid residues: two Pro, two Cys, and one Leu (Table 1). The location of the C-35 ketone (δ_C 212.1) was established based on the presence of a triplet methyl δ_H 1.02 (H-37), a quartet methylene δ_H 2.38 (H-36), and a triplet methylene δ_H 2.36 (H-34) and their corresponding HMBC correlations to the ketone resonance. The combined NMR data were useful in assigning several methylene resonances to the aliphatic chain. However, the precise chain length could not be determined due to the extensive overlap of resonances within the methylene envelope. Fortunately, the molecular formula supported the proposed 14-carbon lipid chain, which is the 12-oxo-tetradecanoyl portion of **1**. Nine of the ten degrees of unsaturation were accounted for with the seven carbonyls and the two Pro, suggesting that a cystine-linked ring was the remaining degree of unsaturation.

The amino acid sequence of eudistomide A (**1**) was determined using a combination of HMBC, ROESY, and MS/MS data. The location of the tetradecanoyl moiety and one of the Cys residues (Cys-2) in **1** was established on the basis of HMBC correlations from the methylenes at δ_H 2.10 and 1.55 (H-25 and H-26, respectively), the Cys-2 α-proton (H-22, δ_H 4.61), and the Cys-2 NH (δ_H 6.08) to a carbonyl at δ_C 171.8 (C-24). The aforementioned methyl ester was assigned to the C-terminal Cys residue (Cys-1) based on an HMBC correlation from the Cys-1 α-proton (H-2, δ_H 4.86) to the methyl ester carbonyl (C-1, δ_C 170.9). The Leu residue was placed adjacent to Cys-1 (Cys-OMe) on the basis of HMBC correlations from the Cys-1 NH (δ_H 7.19) and the Leu β-protons (H-7a, H-7b, δ_H 1.83, 1.48) to the carbonyl at δ_C 171.5 (C-5). Pro-1 was positioned adjacent to the Leu on the basis of HMBC correlations from the Leu NH (δ_H 9.42) and the Pro-1 β-protons (H-13a, H-13b, δ_H 2.35, 2.10) to the carbonyl at δ_C 173.2 (C-11). Because of their overlap, HMBC correlations from the Leu α-proton (H-6, δ_H 4.49) and the Pro-1 α-proton (H-12, δ_H 4.48) could not be utilized. HMBC correlations from the Pro-1 δ-proton (H-15, δ_H 3.56) and the Pro-2 β-protons (H-18a, H-18b, δ_H 2.28, 1.83) to the carbonyl at δ_C 170.1 (C-16) placed Pro-2 adjacent to Pro-1. No HMBC correlations were observed between Pro-2 and Cys-2. ROESY data supported the proposed sequence of the peptide (Figure 1). Although the ROE correlations from δ_H 4.49 (H-6) and δ_H 4.48 (H-12) supported the peptide sequence, the overlap of the proton signals created ambiguity. Therefore, MS/MS studies were conducted to further support the proposed peptide sequence (Table 2). In order to simplify sequence analysis of **1**, the linear peptide desthioeudistomide B (**3**), generated from a Raney Ni desulfurization of **1** (Cys → Ala, ketone → hydroxyl), was analyzed by MS/MS. Compound **3** showed fragment ions consistent with Leu-Ala-OMe, Pro-Leu-Ala-OMe, Pro-Pro-Leu-Ala-OMe, Pro-Leu, and Pro-Pro-Leu partial sequences. Eudistomide A (**1**) showed a fragmentation pattern similar to that of **3** (Pro-Leu and Pro-Pro-Leu were identical), with an addition of 64 Da for the Cys containing fragments, which is consistent with the addition of two sulfurs. On the basis of the HMBC, ROESY, and MS/MS data, the peptide sequence of eudistomide A (**1**) was assigned as cyclized Cys-Pro-Pro-Leu-Cys-OMe, *N*-acylated with a 12-oxo-tetradecanoyl fragment.



It is well-documented that *cis-trans* conformational differences in proline amide bonds can be distinguished in solution

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TABLE 1. NMR Data for Eudistomides A (1) and B (2) (600 MHz, $CDCl_3$)

		eudistomide A (**1**)		eudistomide B (**2**)	
	position	δ_C	δ_H mult (*J*, Hz)	δ_C	δ_H mult (*J*, Hz)[a]
Cys-1 (Cys-OMe)	1	170.9	—	170.9	—
	2	49.9	4.86, ddd (11.0, 9.3, 3.7)	49.9	4.87
	3a	41.5	3.32, dd (14.2, 3.7)	41.5	3.33
	3b		2.77, dd (14.2, 11.0)		2.77
	4	52.5	3.73, s	52.5	3.74
	NH	—	7.20, d (9.3)	—	7.18
Leu	5	171.5	—	171.5	—
	6	51.4	4.49,[b] ddd (8.1, 5.3, 5.0)	51.4	4.50
	7a	36.1	1.83,[b] ddd (14.7, 10.8, 5.0)	36.1	1.83
	7b		1.48,[b] ddd (14.7, 9.3, 5.3)		1.48
	8	24.2	1.60, m (10.8, 9.3, 6.6, 6.4)	24.2	1.60
	9	21.0	0.84, d (6.4)	21.0	0.85
	10	23.0	0.94, d (6.6)	23.0	0.95
	NH	—	9.42, br d (8.1)	—	9.40
Pro-1	11	173.2	—	173.2	—
	12	60.9	4.48,[b] dd (8.2, 2.5)	60.9	4.49
	13a	32.5	2.35,[b] dddd (12.5, 6.5, 6.5, 2.5)	32.5	2.35
	13b		2.10,[b] dddd (12.5, 10.5, 8.2, 1.9)		2.10
	14a	21.3	1.96,[c] m	21.3	1.96
	14b		1.80,[c] m		1.80
	15	46.5	3.56,[b] dd (7.2, 10.0)	46.5	3.56
Pro-2	16	170.1	—	170.1	—
	17	59.4	4.94, dd (8.7, 2.0)	59.4	4.94
	18a	30.7	2.28,[b] dddd (8.1, 8.7, 12.3, 16.4)	30.7	2.28
	18b		1.83,[c] m		1.83
	19a	22.4	1.94,[c] m	22.4	1.94
	19b		1.87,[c] m		1.87
	20a	46.4	3.73,[b] ddd (11.9, 8.8, 4.6)	46.4	3.73
	20b		3.53,[b] ddd (11.9, 7.7, 7.7)		3.53
Cys-2	21	169.4	—	169.4	—
	22	48.8	4.61, ddd (10.5, 10.5, 5.1)	48.8	4.62
	23a	38.4	3.25, dd (12.9, 5.1)	38.4	3.25
	23b		2.56, dd (12.9, 10.5)		2.57
	NH	—	6.08, br d (10.5)	—	5.98
12-oxo-tetradecanoyl	24	171.8	—	171.8	—
	25	36.1	2.10,[c] m	36.1	2.10,[c] m
	26	24.9	1.55,[c] m	24.9	1.55,[c] m
	27−32	29.0	1.23, br s	29.0	1.23, br s
	33a	23.9	1.54,[c] m	25.8	1.38,[c] m
	33b	—	—		1.28, m
	34	42.4	2.36, t (7.5)	36.7	1.37,[c] m
	35	212.1	—	73.1	3.49, m
	36a	35.8	2.38, q (7.3)	29.7	1.47, m
	36b	—	—		1.40, m
	37	7.8	1.02, t (7.3)	10.1	0.92, t (7.5)

[a] Coupling constants for peptide portion are similar to eudistomide A. [b] Obtained from 1D TOCSY. [c] Signals overlapped.

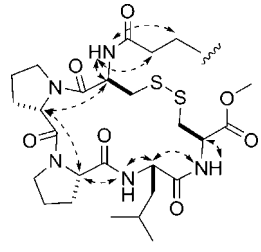

FIGURE 1. Key ROE correlations supporting the peptide sequence of eudistomide A (**1**).

by the chemical shift differential of the β- and γ-carbons ($\Delta\delta_{\beta\gamma}$).[38] Typically, a *cis*-Pro has a chemical shift differential greater than 8 ppm, and a *trans*-Pro has a differential less than 6 ppm. In eudistomide A (**1**), the shift differentials are 11.5 and 8.5 ppm for Pro-1 and Pro-2, respectively, indicating that both Pro residues are in the *cis* conformation.

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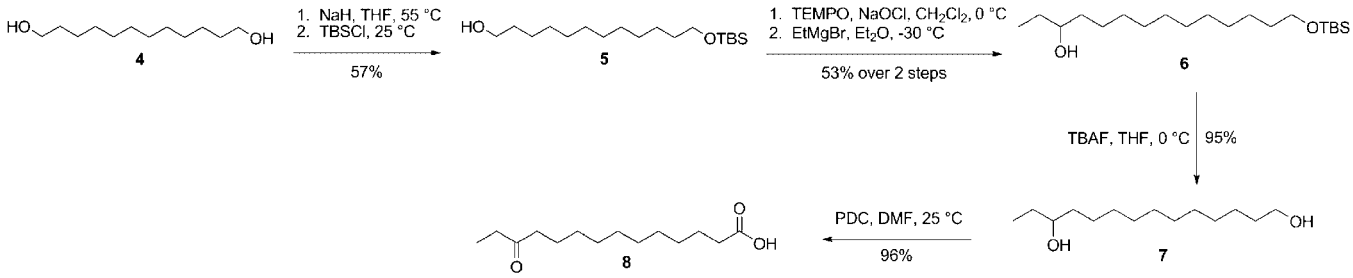
TABLE 2. Fragmentation Ions Observed in ESI-MS/MS Data Confirming the Amino Acid Sequences of Eudistomides A (1) and B (2) and Desthioeudistomide B (3)

compound	ions observed	identity	molecular formula
1, 2	281[a]	Leu-Cys-OMe + S	$C_{10}H_{21}N_2O_3S_2$
1, 2	378[a]	Pro-Leu-Cys-OMe + S	$C_{15}H_{28}N_3O_4S_2$
1, 2	475[a]	Pro-Pro-Leu-Cys-OMe + S	$C_{20}H_{35}N_4O_5S_2$
1, 2	308[a]	Pro-Pro-Leu	$C_{16}H_{26}N_3O_3$
1, 2	211[a]	Pro-Leu	$C_{11}H_{19}N_2O_2$
3	217.15502[b]	Leu-Ala-OMe	$C_{10}H_{21}N_2O_3$
3	314.20796[b]	Pro-Leu-Ala-OMe	$C_{15}H_{28}N_3O_4$
3	411.26097[b]	Pro-Pro-Leu-Ala-OMe	$C_{20}H_{35}N_4O_5$
3	308.19740[b]	Pro-Pro-Leu	$C_{16}H_{26}N_3O_3$
3	211.14442[b]	Pro-Leu	$C_{11}H_{19}N_2O_2$

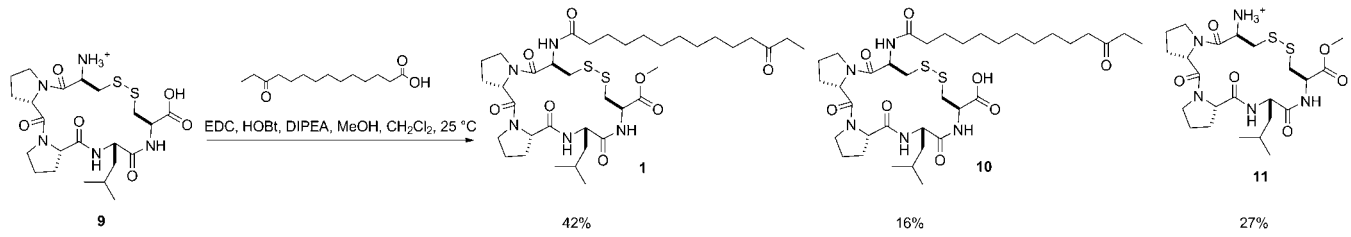
[a] Ions observed on a micro Q-tof using CID. Identical ions observed for synthetic eudistomides A (**1**) and B (**2**). [b] Ions observed on a LTQ-FT using IRMPD; all ions have <1.5 ppm accuracy.

The absolute configuration of each amino acid in eudistomide A (**1**) was determined by chiral HPLC analysis of the acid

SCHEME 1. Synthesis of 12-Oxo-tetradecanoic Acid (8)



SCHEME 2. Synthesis of Eudistomide A (1)



hydrolysate. Compound **1** was desulfurized using Raney Ni in refluxing MeOH[39] to generate desthioeudistomide B (**3**), which was then hydrolyzed with aqueous HCl. Chiral HPLC analysis of both the hydrolysate and authentic standards established the presence of two L-Pro, two L-Ala, and one L-Leu in eudistomide A (**1**). Because the linear peptide **3** contained only L-Ala, the two cysteines in **1** were also assigned L-configuration.

Eudistomide B (**2**) was assigned the molecular formula $C_{37}H_{63}N_5O_8S_2$ on the basis of HRESIMS analysis of the protonated molecular ion $[M + H]^+$ at *m/z* 770.4160. When compared with the formula for **1**, eudistomide B (**2**) showed an additional two H's, which suggested either the disulfide bond or the ketone was reduced in **2**. The NMR spectra of eudistomides A (**1**) and B (**2**) are virtually identical and indicated that both **1** and **2** contained the same five amino acids (Table 1). One obvious difference between the two compounds was the lack of a ketone in **2**. Analysis of COSY and HMBC data showed that a secondary alcohol (δ_C 73.1) was present at C-35 in eudistomide B (**2**). MS/MS fragmentation of **2** confirmed that the disulfide bond was still present (Table 2). Because HMBC, ROESY, and MS/MS data for **2** were comparable to **1**, the peptide sequence of eudistomide A (**2**) was confirmed as cyclized Cys-Pro-Pro-Leu-Cys-OMe, *N*-acylated with a 12-hydroxy-tetradecanoyl fragment. The $\Delta\delta_{\beta\gamma}$ for eudistomide B (**2**) was identical to that for **1**, indicating that **2** also contained two *cis*-Pro residues. Because eudistomides A (**1**) and B (**2**) are biosynthetically related, and all chemical shifts for the peptide portions are virtually indistinguishable, the amino acids in **2** were also assigned the L-configuration.

A total synthesis of eudistomide A (**1**) was undertaken to confirm the proposed structure. To synthesize the 12-oxo-tetradecanoic acid, the commercially available starting material dodecane-1,12-diol (**4**) was protected with TBSCl to give the monoprotected 12-(*tert*-butyldimethylsilyloxy)dodecan-1-ol (**5**) (Scheme 1). Catalytic TEMPO oxidation of compound **5**, followed by Grignard addition of the ethyl group, generated 14-(*tert*-butyldimethylsilyloxy)tetradecan-3-ol (**6**). Compound **6** was deprotected with TBAF to yield tetradecane-1,12-diol (**7**) and subsequently oxidized to form 12-oxo-tetradecanoic acid (**8**). The cyclized pentapeptide, Cys-Pro-Pro-Leu-Cys (**9**), was synthesized by the University of Utah Peptide Synthesis facility using standard solid-phase peptide synthesis procedures. Coupling of the acid (**8**) to the cyclic pentapeptide (**9**) and simultaneous methylation, achieved using EDC, HOBt, DIPEA, and MeOH, yielded eudistomide A (**1**) as the major product. Eudistomide acid (**10**) and the cyclic pentapeptide methyl ester (**11**) were identified as side products (Scheme 2). Interestingly, during the attempt to form the methyl ester (**11**) of precursor peptide (**9**) with diazomethane prior to coupling, the primary amine of Cys-2 was concurrently methylated to form the quaternary amine. Thionyl chloride in MeOH proved to be the best means of introducing the methyl ester to this peptide and can be used prior to coupling without undesired amine methylation. The side products of the coupling reaction (**10** and **11**) were converted to eudistomide A (**1**) in the following manner: Compound **10** was methylated using thionyl chloride in MeOH,[40] and **11** was coupled to the acid (**8**) using EDC, HOBt, and DIPEA. The final synthetic product (**1**) was identical to the natural product obtained from *Eudistoma* sp. in HPLC retention time, MS/MS fragmentation, and MS and NMR spectra, which confirmed the proposed structure for eudistomide A (**1**).

Determining the configuration of the C-35 alcohol in eudistomide B (**2**) was necessary to complete the structure. Several attempts were made to esterify the C-35 alcohol of **2** with MPA to determine the configuration by the modified Mosher method; however, none proved successful. Optimization of reaction conditions for MPA derivitization was carried out using **6** from the synthesis; however, the conditions were not favorable for **2**, and no significant amounts of products were detected by LC-MS. To preserve the limited supply of eudistomide B (**2**), a scheme using synthetic eudistomide A (**1**) was proposed in an attempt to resolve the configuration of the C-35 alcohol in **2** (Scheme 3). Compound **1** was reduced using sodium borohy-

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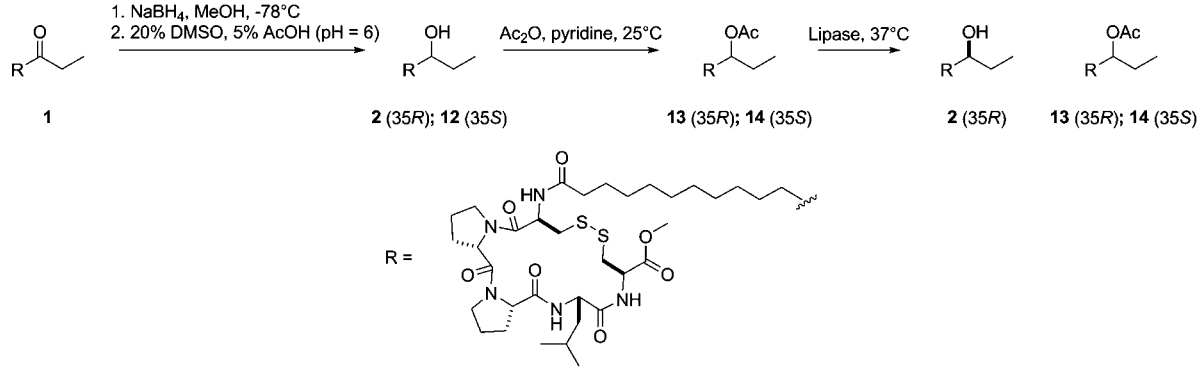
SCHEME 3. Determination of the Side Chain Configuration in Eudistomide B (2)



dride in MeOH at −78 °C,[41] and the disulfide was reoxidized using DMSO in acetic acid[42] to give a mixture of C-35 alcohol epimers (**2**, **12**). The HSQC data showed that the ^{1}H and ^{13}C chemical shifts of the C-35 hydroxy methines of the two epimers were very different; one showed a proton resonance at δ_H 3.52 attached to a carbon at δ_C 73.2, and the other showed a proton resonance at δ_H 4.84 attached to a carbon at δ_C 74.7. Efforts to separate the alcohol epimers (**2**, **12**) were not successful. Literature precedents using similar substrates[43,44] confirm that lipase B from *Candida antarctica* shows selectivity for *R*-secondary acetates as substrates, with % ee being ≥99% when conversion is less than 50%. Accordingly, lipase-catalyzed hydrolysis of the C-35 acetoxy epimers would primarily generate the *R*-alcohol. Therefore, the alcohol epimers were treated with acetic anhydride in pyridine to generate the C-35 acetoxy derivatives (**13**, **14**). Lipase B from *C. antarctica* was added to the acetoxy epimers (**13**, **14**) at 37 °C, and the products were analyzed by HSQC. Only one alcohol signal appeared in the HSQC (δ_H 3.52, δ_C 73.2) and was assigned the *R*-configuration. The C-35 alcohol in eudistomide B (**2**) (δ_H 3.49, δ_C 73.1) also was assigned the *R*-configuration on the basis of identical chemical shifts to the lipase-derived *R*-isomer of **1**.

Eudistomides A (**1**) and B (**2**) are interesting cyclic lipopeptides that contain several rare structural motifs. The acyl chain present in **1** and **2** has never been reported from a marine organism.[45] However, similar acyl groups, such as unsubstituted tetradecanoyl,[46−48] 13-methyltetradecanoyl,[49,50] 2,3-hydroxytetradecanoyl,[51] 3-hydroxy-13-methyltetradecanoyl,[52] and 7-methoxytetradec-4-enoyl,[53−55] have been reported. Interestingly, the microsporins[56] contain a 2-amino-8-oxodecanoic acid or a 2-amino-8-hydroxydecanoic acid that is similar to the ketone and hydroxyl derivatives seen in **1** and **2**. While marine organisms are the source of several peptides that contain cystine moieties such as the ulithiacyclamides,[1−3] thiocoralines,[57] microcionamides,[39] and neopetrosiamides,[58] eudistomides A (**1**) and B (**2**) are the first ascidian-derived peptides cyclized solely by a disulfide bridge.[45]

Experimental Section

Biological Material. *Eudistoma* sp., sample FJ04-12-071, was collected by SCUBA near Namena Barrier Reef, Fiji Islands (17° 06.884′ S, 179° 03.805′ E); a voucher specimen is maintained at the University of Utah. This thick, dark gray, encrusting *Eudistoma* sp. was found in large patches (approximately 0.5 m) and expressed copious amounts of clear mucous.

Extraction and Isolation. The *Eudistoma* sp. specimen was lyophilized and ground to a fine powder. The powder was exhaustively extracted with MeOH to yield 15.4 g of crude extract. The crude extract was subjected to an EtOAc/H_2O partition to generate 1.7 g of the organic fraction. The EtOAc soluble material was separated on HP20SS resin using a gradient of H_2O to acetone in 10% steps and a final wash of 100% acetone to yield 11 fractions. The sixth (50/50 acetone/H_2O) and seventh fractions (60/40 acetone/H_2O) were combined (34.5 mg) and chromatographed by HPLC using a Phenomenex Luna C_{18} column (250 × 10 mm) employing a gradient of 50% CH_3CN/H_2O to 100% CH_3CN at 2.5 mL/min over 30 min to yield four fractions, 29A−29D. Fraction 29C was further chromatographed using a Phenomenex Luna C_{18} (150 × 4.6 mm) column using a gradient of 70% CH_3CN/H_2O to 85% CH_3CN/H_2O at 1.0 mL/min over 15 min to yield two fractions, 35A and 35B. Fraction 35A was repurified using a Phenomenex Luna C_{18} (250 × 10 mm) column using a gradient of 50% CH_3CN/50% H_2O (+0.1% AcOH) to 98% CH_3CN/2% H_2O (+0.1% AcOH) at 4.5 mL/min over 20 min to yield eudistomide A (**1**, 0.2 mg) eluting at 12.9 min, and eudistomide B (**2**, 0.35 mg) eluting at 11.2 min. Reisolation from associated fractions and extraction of the remaining crude material provided an additional 3.2 mg of **1**.

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Eudistomide A (1): amorphous, white solid; $[\alpha]^{20}_D$ −1.3 (*c* 0.2, MeOH); UV (MeOH) λ_{max} (log ε) 206 (3.77) nm; IR (film, NaCl) ν_{max} 3284 (br), 2921, 2852, 1743, 1708, 1662, 1641, 1549, 1253, 702 cm^{-1}; ^1H and ^{13}C NMR data (Table 1); HRESIMS *m/z* 790.3897 $[M + Na]^+$ (calcd for $C_{37}H_{61}N_5O_8S_2Na$, 790.3859).

Eudistomide B (2): amorphous, white solid; $[\alpha]^{20}_D$ −4.0 (*c* 0.03, MeOH); UV (MeOH) λ_{max} (log ε) 206 (3.61) nm; IR (film, NaCl) ν_{max} 3222 (br), 2921, 2852, 1745, 1662, 1641, 1549, 1259, 725 cm^{-1}; ^1H and ^{13}C NMR data (Table 1); HRESIMS *m/z* 770.4160 $[M + H]^+$ (calcd for $C_{37}H_{64}N_5O_8S_2$, 770.4191).

Desulfurization of Eudistomide A (1). Approximately 100 μL of Raney 2800 nickel (50% slurry in H_2O, excess) was added to eudistomide A (1.0 mg, 1.3 μmol) in MeOH (1.2 mL). Argon was bubbled through the solution to remove O_2; the resulting black suspension was heated at 65 °C under argon for 4 h, and monitored by HPLC for the disappearance of starting material. Upon cooling, the solution was purified on a C_{18} SPE cartridge using MeOH as eluant, yielding pure desthioeudistomide B (**3**, 0.5 mg, 54% yield).

Desthioeudistomide B (3): amorphous, white solid; HRESIMS *m/z* 708.49059 $[M + H]^+$ (calcd for $C_{37}H_{66}N_5O_8$, 708.49059); HRESI-MS/MS (Table 2).

Absolute Configuration of Each Amino Acid in Desthioeudistomide B (3). Compound **3** (0.5 mg) was dissolved in 6 N HCl (500 μL) and heated at 110 °C under argon for 21 h. The product mixture was lyophilized and analyzed by chiral HPLC [column, Phenomenex Chirex phase 3126 (D) (250 × 4.6 mm); solvent 1 mM $CuSO_4/CH_3CN$ (95/5); flow rate, 1.0 mL/min; UV detection at 254 nm], comparing the retention times to those of authentic standards. The retention times (min) of the authentic amino acids were 6.9 for L-Ala, 8.4 for D-Ala, 9.5 for L-Pro, 20.0 for D-Pro, 51.3 for L-Leu, and 65.9 for D-Leu. The retention times of the amino acid components in the acid hydrolysate were 6.9, 9.4, and 51.3, indicating the presence of L-Ala, L-Pro, and L-Leu, respectively.

12-(*tert*-Butyldimethylsilyloxy)dodecan-1-ol (5). A total of 0.4 g of NaH (60% dispersion in mineral oil, 10 mmol) was washed with anhydrous THF (2 × 20 mL, to remove the mineral oil) and suspended in anhydrous THF (20 mL). A total of 2.02 g (10 mmol) of dodecane-1,12-diol (**4**) was added to the solution, and the solution was left to stir under argon at 55 °C. After 18 h, the mixture was cooled to rt, and 1.51 g of TBSCl (10 mmol) in anhydrous THF (2.5 mL) was added and allowed to stir for 2 h at rt. The mixture was diluted with Et_2O and washed successively with 10% K_2CO_3 and brine. The combined organic extracts were dried ($MgSO_4$) and concentrated in vacuo. Purification of the crude material by silica column chromatography [solvent 1, 90% hexanes/10% EtOAc (elutes the disilyl ether product); solvent 2, 80% hexanes/20% EtOAc] afforded the monosilylated product **5** as a colorless solid (1.01 g, 57% yield). **5**: ^1H NMR (CD_3OD, 400 MHz) δ 3.57 (2H, t, J = 6.4 Hz), 3.48 (2H, t, J = 6.7 Hz), 1.45 (4H, m), 1.28 (14H, br s), 0.84 (9H, s), 0.00 (6H, s); ^{13}C NMR (CD_3OD, 400 MHz) δ 63.4, 63.1, 34.0, 33.8, 31.0−30.5, 27.1, 27.0, 26.6, 19.3, −5.0; HRESIMS *m/z* 299.2765 $[M - H_2O]^+$ (calcd for $C_{18}H_{39}OSi$, 299.2770).

14-(*tert*-Butyldimethylsilyloxy)tetradecan-3-ol (6). A 0.5 M aq solution of KBr (7.14 mg, 0.06 mmol) was added to a vigorously stirring solution of **5** (188.8 mg, 0.6 mmol) in CH_2Cl_2 (0.7 mL) at 0 °C, followed by the addition of the TEMPO (2,2,6,6-tetramethylpiperdine 1-oxyl) free radical (0.94 mg, 0.006 mmol). NaOCl (5% active chlorine), adjusted to pH 8.6 using $NaHCO_3$ (1.5 mL), was added dropwise to the solution; the mixture was allowed to stir for 5 min at 0 °C until the hypochlorite was consumed, and the solution went from a red-orange to a milky yellow color. The organic layer of the two-phase mixture was separated, dried ($MgSO_4$), and concentrated to dryness in vacuo at 0 °C. The crude aldehyde (160 mg, 0.51 mmol) was dissolved in anhydrous Et_2O (3.2 mL) and cooled to −30 °C under argon. EtMgBr (202 μL, 0.61 mmol, 3 M in ether) was added dropwise to the solution, and the mixture was left to stir at −30 °C. After 1.5 h, the reaction was quenched with saturated NH_4Cl (316 μL). The solution was warmed to rt, washed with brine, and extracted with Et_2O. The ethereal extracts were combined, dried ($MgSO_4$), filtered, and concentrated in vacuo. The crude product was purified by silica column chromatography (solvent, 83% hexanes/17% MTBE) to afford **6** as a colorless solid (91.8 mg, 53% yield). **6**: ^1H NMR ($CDCl_3$, 400 MHz) δ 3.55 (2H, t, J = 6.7 Hz), 3.47 (1H, m), 1.55−1.30 (6H, m), 1.24 (16H, br s), 0.89 (3H, t, J = 7.5 Hz), 0.84 (9H, s), 0.00 (6H, s); ^{13}C NMR ($CDCl_3$, 400 MHz) δ 73.5, 63.6, 37.2, 33.1, 30.4, 30.0−29.5, 26.2, 26.0, 25.9, 18.6, 10.1, −5.0; HRESIMS *m/z* 367.3009 (calcd for $C_{20}H_{44}O_2SiNa$, 367.3008).

Tetradecane-1,12-diol (7). TBAF (tetrabutylammonium fluoride, 124.5 mg, 0.48 mmol) in THF (0.5 mL) was added dropwise to a solution of **6** (81.8 mg, 0.24 mmol) in THF (3.1 mL) at 0 °C. The reaction mixture was warmed to rt and stirred for 4 h. Cold water was added to quench the reaction, and the resultant mixture was extracted with EtOAc. The combined organic extracts were washed with brine, dried ($MgSO_4$), filtered, and concentrated to dryness in vacuo. The crude product was purified by silica column chromatography [solvent 1, 80% CH_2Cl_2/20% EtOAc (eluted remaining **6**); solvent 2, 50% CH_2Cl_2/50% EtOAc] to yield **7** as a colorless solid (52.0 mg, 95% yield). **7**: ^1H NMR ($CDCl_3$, 400 MHz) δ 3.58 (2H, t, J = 6.6 Hz), 3.46 (1H, m), 1.60−1.30 (8H, m), 1.24 (14H, br s), 0.89 (3H, t, J = 7.4 Hz); ^{13}C NMR ($CDCl_3$, 400 MHz) δ 73.5, 63.2, 37.1, 33.0, 30.3, 30.0−29.5, 25.9, 25.8, 10.1; HRESIMS *m/z* 213.2217 $[M - H_2O]^+$ (calcd for $C_{14}H_{29}O$, 213.2218).

12-Oxo-tetradecanoic Acid (8). A solution of **7** (15 mg, 65.2 μmol) in anhydrous DMF (700 μL) was treated with PDC (197 mg, 0.52 mmol) under argon. The dark brown solution was allowed to stir at rt for 17 h. The reaction was quenched with water and extracted with EtOAc. The combined organic layers were washed successively with water and brine, dried ($MgSO_4$), and concentrated to dryness in vacuo to yield **8** as a colorless solid (15.1 mg, 96% yield). **8**: ^1H NMR ($CDCl_3$, 400 MHz) 2.50−2.20 (6H, m), 1.60−1.40 (4H, m), 1.23 (12H, br s) 1.01 (3H, br t, J = 6.5 Hz); ^{13}C NMR ($CDCl_3$, 400 MHz) δ 212.0, 179.7, 42.3, 35.7, 33.9, 29.5−28.5, 24.5, 23.8, 7.7; HRESIMS *m/z* 243.1946 $[M + H]^+$ (calcd for $C_{14}H_{27}O_3$, 243.1955).

Cyclic pentapeptide (9): amorphous, white solid; ^1H and ^{13}C NMR data (Supporting Information); HRESIMS *m/z* 552.1912 $[M + Na]^+$ (calcd for $C_{22}H_{35}N_5O_6S_2Na$, 552.1926).

Synthesis of Eudistomide A (1). EDC (12.6 mg, 22.7 μmol) and HOBt (8.9 mg, 22.7 μmol) were added to a solution of **8** (5.1 mg, 22.7 μmol) in anhydrous 90% CH_2Cl_2/10% MeOH (1.8 mL) under argon and allowed to stir for 3 h to activate the acid. DIPEA (3.3 μL, 19.3 μmol) was added to a stirred solution of **9** (10.2 mg, 19.3 μmol) in 85% CH_2Cl_2/15% MeOH (1.9 mL) and allowed to stir for 3 h to neutralize the peptide. After 3 h, the neutralized peptide (**9**) was added to the acid (**8**), and DIPEA was added to the solution (3.3 μL, 19.3 μmol). The reaction was monitored by HPLC for the disappearance of starting material. The reaction was quenched with H_2O after 48 h, extracted with CH_2Cl_2, and concentrated in vacuo. The reaction products were purified using a Phenomenex C_{18} (250 × 10 mm) column using a gradient of 2% CH_3CN/98% H_2O (+0.1% AcOH) to 98% CH_3CN/2% H_2O (+0.1% AcOH) at 4.5 mL/min over 40 min to yield pure eudistomide A (**1**) (2.2 mg, 42%) eluting at 33.4 min, the peptide methyl ester (**11**) (1.0 mg, 27%) eluting at 11.2 min, and the eudistomide acid (**10**) (0.8 mg, 16%) eluting at 28.3 min. HPLC retention times of the natural eudistomide A (**1**) and the synthetic eudistomide A (**1**) were compared using a Phenomenex C_{18} (250 × 4.6 mm) column employing a gradient of 50% CH_3CN/50% H_2O (+0.1% AcOH) to 98% CH_3CN/2% H_2O (+0.1% AcOH) at 1 mL/min over 20 min. Natural **1** had a retention time of 12.8 min, and synthetic **1** also had a retention time of 12.8 min. **1**: $[\alpha]^{20}_D$ −3.5 (*c* 0.06, MeOH); ^1H and ^{13}C NMR data identical to natural product (Table 1); HRESIMS *m/z* 768.4034 $[M + H]^+$ (calcd for $C_{37}H_{62}N_5O_8S_2$, 768.4040); ESI-MS/MS (Table 2).

Eudistomide B Epimers (2, 12). A solution of **1** (1.3 mg, 1.7 μmol) in anhydrous MeOH (250 μL) at −78 °C was treated with

$NaBH_4$ (1.0 mg, 26.4 μmol) under argon. The solution was allowed to stir for 7 h. The reaction was quenched with water, the MeOH was evaporated under argon, and the resulting aqueous layer was extracted with CH_2Cl_2. The combined organic extracts were dried under argon and redissolved in 5% AcOH (400 μL, pH 6). DMSO (100 μL) was added to the solution that was left to stir overnight. The reaction products were concentrated in vacuo and purified using a Phenomenex C_{18} (250 × 10 mm) column employing a gradient of 50% CH_3CN/50% H_2O (+0.1% AcOH) to 98% CH_3CN/2% H_2O (+0.1% AcOH) at 4.5 mL/min over 20 min to yield a mixture of C-35 alcohol epimers (**2**, **12**) coeluting at 11.3 min (1.1 mg, 84%). HPLC retention times of the natural eudistomide B (**2**) and the synthetic C-35 alcohol epimers (**2, 12**) were compared using a Phenomenex C_{18} (250 × 4.6 mm) column employing a gradient of 50% CH_3CN/50% H_2O (+0.1% AcOH) to 98% CH_3CN/2% H_2O (+0.1% AcOH) at 1 mL/min over 20 min. Natural **2** had a retention time of 11.2 min, and the synthetic alcohol epimers (**2**, **12**) also had retention times of 11.2 min. **2**, **12**: LRESIMS *m*/*z* 770.4 $[M + H]^+$; ESI-MS/MS (Table 2).

Acknowledgment. The authors wish to acknowledge Dr. Krishna Parsawar and Dr. Chad Nelson, University of Utah Mass Spectrometry and Proteomics Core Facility, for performing FTMS experiments. Ascidian taxonomy was graciously provided by Gretchen Lambert, University of Washington, Friday Harbor Laboratories. The authors also would like to recognize Dr. Robert Schackmann and Scott Endicott of the University of Utah Peptide Synthesis Core Facility. We acknowledge the NIH, Grants RR13030, RR06262, and RR14768, and NSF Grant DBI-0002806 for funding NMR instrumentation in the University of Utah Health Sciences NMR Facility. This research was supported by NCI Grant CA67786 and NIH Grant CA36622.

Note Added after ASAP Publication. There were numbering errors in Schemes 1−3 in the version published ASAP December 3, 2008; the revised version was published ASAP December 11, 2008.

Supporting Information Available: General experimental procedures, acetylation reaction conditions, lipase reaction conditions, ^1H spectra of **1** and **2**, ^{13}C spectrum of **1**, ^1H and ^{13}C spectra of **5**−**9**, table of assignments for **9**, ^1H and HSQC spectra of eudistomide B epimers (**2**, **12**), ^1H and HSQC spectra of eudistomide B acetate ester epimers (**13**, **14**), and ^1H and HSQC spectra of the lipase reaction products (**2**, **13**, **14**). This material is available free of charge via the Internet at http://pubs.acs.org.

JO8022582